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Research Article

Mortality Patterns in Critically III Patients on Mechanical Ventilation with Candida Colonization of the Respiratory Tract with or without Gram-Negative Bacteria Ventilator-Associated Pneumonia in a Tertiary Hospital in Saudi Arabia - a

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ABSTRACT

Purpose: Candida is often regarded as being non-pathogenic. Recent studies suggest that a high mortality exists among patients who have Candida and Gram-Negative Bacteria (GNB) Ventilator-Associated Pneumonia (VAP). This study determined mortality levels among patients with bronchial Candida colonization with or without GNB VAP. Mortality patterns between Candida albicans and non-albicans were also determined.

Materials and Methods: Critically ill patients on mechanical ventilation (MV) \ge 48 hours enrolled from January to December 2019 were classified into four main groups based on tracheal aspirate culture (C. albicans only, C. albicans + GNB, C. non-albicans only and C. non-albicans + GNB.

Results: Mortality among the 173 patients was high and ranged from 50.0 % with C. albicans + GNB to 77.4 % with C. non-albicans + GNB. No significant difference in mortality occurred between compared groups except C. albicans + GNB versus C. non-albicans + GNB where a statistically significant odds ratio was found (2.73; 95 % confidence interval, 1.03-7.5; p = 0.025).

Conclusions: There is high mortality with Candida species colonization especially with concomitant GNB VAP. The highest mortality is with C. non-albicans. Further studies are needed to confirm this finding.

Keywords: Candida colonization; Mechanical ventilator; Ventilator-associated pneumonia; Gram-negative bacteria; Intensive care unit

INTRODUCTION

There has been an increase in nosocomial fungal infections in the past two decades. This is considered problematic especially among Intensive Care Unit (ICU) patients [1]. Candida is an opportunistic pathogen that is typically observed among critically ill patients with compromising underlying conditions. This includes patients who endured multiple invasive procedures or patients who experienced prolonged hospitalization periods for various reasons including multiorgan failure [2]. Other factors such as diabetes mellitus, immunosuppressive therapy (including chemotherapy), and the overuse of extended spectrum antibiotics increased the risk for infections [3].

Candida species are frequently recovered from respiratory tract secretions in mechanically ventilated ICU patients. From a cohort study of 1,107 ICU patients with a median length of hospital stay of 17 days, 834 (75.3 %) were in the Candida species colonization group [4]. In another Canadian study done on ICU patients, 50 % of the patients grew positive Candida on respiratory or rectal cultures. The different Candida species from the respiratory or rectal cultures included *C. albicans* (72 %), *C. glabrata* (16 %), *C. tropicalis* (5 %), and *C. parapsilosis* (3 %) [5].

Of all Candida species, Candida albicans is the most pathogenic and most frequently identified in various candidiasis lesions in humans [6]. Over the last two decades, C. albicans was the most (80 %) recovered Candida species from patients with oral and systemic candidiasis. However, over the same period, the number of infections caused by non-albicans species has increased significantly [7].

Bronchial Candida species colonization is a common condition among critically ill patients [8,9]. Although Candida species are considered to be non-invasive and not usually implicated in pneumonia, in recent studies, there have been conflicting results [8-10]. A significant association between the presence of Candida species in the respiratory tract of patients and hospital mortality has been reported [11]. Candida colonization of the respiratory tract has also been implicated with bacteria Ventilator-Associated Pneumonia (VAP) [11]. Delislee, et al. [11] hypothesized that the presence of Candida species in respiratory tract secretions may explain the excess morbidity and mortality seen in critically ill patients with or without positive bacterial culture. Their conclusion suggests that the presence of Candida species in critically ill patients may be a pointer to the severity of disease. However, there is inadequate evidence on the hospital mortality among critically ill patients on Mechanical Ventilator (MV) with Candida in Saudi Arabia. Thus, the primary objective of this study was to determine the hospital mortality among critically ill patients on MV with respiratory tract culture of Candida species, with or without the presence of Gram-Negative Bacteria (GNB) and to determine whether mortality would differ between Candida albicans and non-albicans.

MATERIALS AND METHODS

Study setting

Approval for this study was obtained from the ethics committee of King Abdul-Aziz University Hospital and the Ministry of Education. This study was conducted at the Molecular and Clinical Microbiology Laboratory at a teaching tertiary hospital, Jeddah, kingdom of Saudi Arabia from January 2019 till December 2019.

Study design

Patients: Patients who met the inclusion criteria were 173 critically ill patients who had been on MV for 48 hours or more. We excluded patients with fungemia (defined as the presence of fungi or yeasts in the blood) and patients on antifungal treatment The patients were classified into four main groups based on the presence of Candida species with or without GNB VAP in the patients' tracheal aspirate as follows: *C. albicans* alone group or *C. albicans* and GNB group or *C. non-albicans* and GNB group

Trachea aspirate culture procedure

a. Identification and susceptibility of bacteria isolated from tracheal aspirate samples: The trachea aspirate samples obtained from each patient were cultured for bacteria and fungi.

Each tracheal aspirate specimen was processed and cultured on Blood agar, MacConkey agar and Chocolate agar and Sabouraud Agar (SDA: Saudi Prepared Media Laboratories, Riyadh, Kingdom of Saudi Arabia). The cultured plates were incubated for 18 to 24 hours (MacConkey agar at 35-37°c in ordinary incubator (Forma



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Scientific Incubator, Germany). Blood agar and Chocolate agar plates were incubated at 35-37°c in 5-10 % CO2 incubator (Sanyo CO2 Incubator, Japan). The colonies were identified by gram staining for further identification and Antibiotic Sensitivity Testing (AST) of bacteria by Vitek * 2 system (bioMerieux, Inc., France). The isolated pure colonies were selected and a purity plate was done to ensure that a pure culture was used for testing. Then, 3 ml of 0.45 % sterile saline was aseptically added into clear plastic test tube. A sufficient number of morphologically similar colonies were then transferred by a sterile loop to the saline tube and its density was checked by using Vitek 2 DensiCheck (bioMerieux, Inc., France) equivalent to 0.5 to 0.63 McFarland. The suspension tube was placed in the cassette, followed by an empty tube. The identification card was placed in the suspension tube and the AST card was placed in the empty tube. When the sample cycle was finished, the cassettes and the tubes were discarded. Minimal Inhibitory Concentration (MIC) was calculated and represented as (sensitive, intermediate or resistant). MIC of amikacin and meropenem with Acinetobacter baumannii were not done by Vitek 2 system, but done by E- test strips (bioMerieux SA, RCS LYON, Marcy-l' Etolle, France).

The GNB identified included Pseudomonas aeruginosa, Klebsiella pneumoniae, Escherichia coli, Proteus mirabilis, Serratia marcescens, Enterobacter cloacae, Enterobacter aerogenes, Stenotrophomonas maltophila, and Haemophilus influenza. Of the GNB, 37(60.7 %) were Multi-Drug Resistant (MDR).

b. Identification of yeast isolated from tracheal aspirate samples by API°20 C AUX: (bioMerieux, Inc., France)

A portion of yeast colonies was picked up from a young culture (18-24 h) on SDA and a suspension with a turbidity equal to 2 McFarland was prepared. Then 100 µl of this suspension was transferred to the ampule of the API C medium and homogenized by a pipette; they were then added into the cupules of each strip that contained the API suspension medium. The strip was incubated at $29^{\circ}C \pm 2^{\circ}C$ for 48-72 hours and after that, the identification was done automatically by the ATB^{**}/Mini API^{*} instrument (bioMerieux SA, Marcy-l' Etolle, France) using database (V4.0).

Data collection

Patients' baseline characteristics, including age and underlying conditions as well as the baseline laboratory parameters were obtained from the patients' clinical records. The length of ICU stay, number of days on mechanical ventilation before culture and the number of days on prior anti-microbial treatment were recorded for each patient. To determine disease severity at ICU admission, the Acute Physiologic Assessment and Chronic Health Evaluation (APACHE) scoring system was used.

Statistical analysis

Data were analyzed using Statistical Package for the Social Sciences (SPSS) version 18 software. Continuous variables were presented as mean, standard deviation, and range. Categorical variables were reported in percentages. Chi-square test was utilized to test for the association and/or difference between categorical variables. Mortality outcomes among the C. albicans and nonalbicans groups were reported as percentages and compared using chi square test. Mortality differences between C. albicans and nonalbicans groups were determined between the following groups: C. albicans only versus C. albicans + GNB, non-albicans only versus non-albicans + GNB, C. albicans only versus non-albicans only as well as C. albicans + GNB versus non-albicans + GNB. Odds ratio and 95 % confidence interval were reported. P-values less than 0.05 were considered statistically significant.

RESULTS

Patients' demographic and clinical characteristics

Overall, 173 patients met the inclusion criteria and were enrolled from January 2019 till December 2019. The baseline demographic and clinical characteristics are shown in Table 1. The overall age ranged from 17 to 96 years and the majority, 105 (60.7 %) were males. Most common underlying diseases included diabetes mellitus, 82 (47.4 %) followed by heart diseases, 66 (38.2 %), length of ICU stay and APACHE score was similar among all groups as shown in table 1.

Mortality outcomes among C. albicans and non-albicans groups

In table 2, mortality was least with 50.0 % among the C. albicans + GNB group while the highest mortality occurred with 77.4 % among the C. non-albicans + GNB group. More than half (55.9 %) and (58.5 %) mortality occurred among the *C. albicans* and the *C. non-albicans* only groups respectively. There were no significant differences

	C. albicans alone group	C. albicans and GNB group	Non - albicans alone group	Non - albicans and GNB group
	n= 59 (%)	n= 30 (%)	n= 53 (%)	n= 31 (%)
Age	(17 - 90)	(17 - 96)	(15 - 89)	(23 - 84)
Range Mean ± SD	56.13 ± 19.29	60.37 ± 17.05	57.39 ± 18.8	60.96 ± 15.52
Sex				
Male	27 (45.8 %)	22 (73.3 %)	35 (66 %)	21 (67.6 %
Female	32 (54.2 %)	8 (26.7 %)	18 (34 %)	10 (32.3 %
Underlying diseases COPD	3 (5.2 %)	1 (3.3 %)	1 (1.9 %)	3 (9.7 %)
Chronic renal impairment	13 (22 %)	8 (26.7 %)	14 (26.4 %)	9 (29 %)
Heart diseases	22 (37.3 %)	12 (40 %)	21 (39.6 %)	11 (35.5 %
Liver diseases	2 (3.4 %)	1 (3.3 %)	1 (1.9 %)	2 (6.5 %)
Surgical operations	13 (22 %)	1 (3.3 %)	7 (13.2 %)	1 (3.2 %)
Diabetes Millitis	21 (35.6	15 (50 %)	26 (49.1 %)	20 (64.5 %
Immunosuppression	10 (16.9 %)	1 (3.3 %)	3 (5.7 %)	4 (12.9 %
Stroke	5 (8.5 %)	1 (3.3 %)	4 (7.5 %)	0 (0 %)
No of days of ICU stay	4.57 ± 5.46	10.16 ± 21.5	3.56 ± 5.05	6.9 ± 8.59
Mechanical ventilation before culture	50 (84.1 %)	20 (66.6 %)	32 (60.4 %)	17 (54.8 %
Mean ± SD (days)	4.2 ± 5.53	9.9 ± 21.6	3.13 ± 5.26	6.38 ± 8.8
Prior anti - microbial treatment	55 (93.2 %)	25 (83.3 %)	34 (64.2 %)	21 (67.7 %
Mean ± SD (days)	8.13 ± 11.88	10.76 ± 19.80	4.24 ± 6.23	7.58 ± 10.16
Apache score	9–32	12–33	13–31	17–34
Range Mean ± SD	20.74 ± 6.16	20.86 ± 4.19	20.47 ± 4.61	23.24 ± 4.

American J Anesth Clin Res

	Deceased n=103 n (%)	Survived n=70 n (%)	X²	P - value	95 % CI	Mean ± SD of day till death	28 days Mortality n (%)
C. albicans alone group n= 59	33 (55.9)	26 (44.1)	0.28	0.59	0.80 (0.40 - 1.59)	15.24 ± 10.65	30 (90.9)
C. albicans and GNB Group n= 30	15 (50)	15 (50)	0.99	0.31	0.62(0.26 - 1.46)	25.66 ± 23.67	10 (66.7)
C. non - albicans alone group n= 53	31 (58.5)	22 (41.5)	0.03	0.85	0.90(0.42 - 1.84)	13.35 ± 13.73	26 (83.9)
C. non - albicans and GNB n= 31	24 (77.4)	7 (22.6)	4.98	0.025*	2.73(1.03 - 7.5)	13.54 ± 10.55	20 (83.3)

between those who died and those who survived with *C. albicans* only, Odds Ratio (OR), 0.80; 95 % Confidence Interval [CI], 0.4-1.59; p = 0.59, *C. albicans* + GNB OR, 0.62; 95 % CI, 0.26-1.46; p = 0.31) as well as non-albicans only OR, 0.90; 95 % CI, 0.42-1.84; p = 0.85. However, the chi square test showed a significant difference between those who died and those who survived with C. non-albicans + GNB OR, 2.73; 95 % CI, 1.03-7.5; p = 0.025

Mortality differences between *C. albicans* and non-albicans groups with χ^2 and *p*-value

In table 3, the findings of the comparison between groups are shown. No significant differences occurred when *C. albicans* only versus *C. albicans* + GNB or non-albicans only versus non-albicans + GNB or *C. albicans* only versus non-albicans only were compared. However, the comparison of mortality between only two groups (C. albicans + GNB versus C. non-albicans + GNB) showed statistically significant difference (p < 0.05).

DISCUSSION

Candida colonization remains a dilemma among mechanically ventilated patients. The primary objective of our study is to determine the mortality among ventilated patients with Candida colonization, with or without the presence of GNB and to determine mortality differences among different Candida species groups.

Based on the four groups of patients studied, we observed high mortality with Candida species colonization with *C. albicans* only group and the non-albicans only group (55.9 % and 58.5 % respectively). This same finding has been observed in other studies. Hamet, et al. [12] found that mortality was significantly higher in mechanically ventilated patients colonized with Candida (44 %) versus controls (31 %) [12]. Other studies showed similar higher mortality findings among patients with Candida colonization (43 % and 34 %) and controls (36 % and 21 %) by Azoulay, et al. [10] and Delisle, et al. [13] respectively. The consistency of findings persisted over the years as reported in 2011 by Delisle, et al. [11], among the same patient population with suspected VAP and no bacterial culture growth, but with Candida colonization, higher mortality even after controlling and adjusting for confounders persisted.

We found a very high mortality with Candida non-albicans + GNB in our study. From the findings of a few studies on the virulence

Table 3: Mortality differences between C. albicans and non - albicans groups with χ^2 and p – value.

	Deceased Survived			
	n=103 (%)	n=70 (%)	X ²	P value
Albicans alone (n=59)	33 (55.9 %)	26 (44.1 %)	0.28	0.99
Albicans + GNB (n=30)	15 (50 %)	15 (50 %)		
Non albicans alone (n=53)	31 (58.5 %)	22 (41.5 %)	3.1	0.78
Non albicans + GNB (n=31)	24 (77.4 %)	7 (22.6 %)		
Albicans alone (n=59)	33 (55.9 %)	26 (44.1 %)	0.07	0.78
Non albicans alone (n=53)	31 (58.5 %)	22 (41.5 %)		
Albicans + GNB (n=30)	15 (50 %)	15 (50 %)	4.97	0.025*
Non albicans + GNB (n=31)	24 (77.4 %)	7 (22.6 %)		

*p < 0.05; χ^2 Chi square statistical test; GNB: Gram-Negative Bacteria; n: Sample Size.

factors of non-albicans, the mortality differences observed in our study among Candida species may be explained by their virulence factors. Biofilms is one of the virulence factors of Candida species. Deorukhkar, et al. [14] noted a greater biofilm forming ability in *C. tropicalis* compared to *C. albicans* while Pongracz, et al. [15] reported that C. non-albicans produce more biofilm than C. albicans. Biofilm formation protects Candida species against host defenses and also carries a significant resistance to antifungal therapy [16]. Significant antifungal activities have been reported in a study [17].

There is a high mortality observed in our study from the combination of fungi and bacteria. Evidence from previous studies showed that *C. albicans* protects bacteria from clearance by the host defense system and also enhances bacterial virulence [18]. There is a strong interaction between Candida, gram-positive bacteria and GNB through quorum sensing which explains the protective role that the Candida plays to the bacteria [19]. This might explain our finding of a high mortality with the combination of fungi and bacteria. It has also been demonstrated in many studies that Candida species colonization increases inflammation and is associated with poor clinical outcome [20-22]. However, similar findings have also been observed with other GNB VAP such as Pseudomonas. Azoulay, et al. [10] studied

American J Anesth Clin Res

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colonized and non-colonized Candida in mechanically ventilated patients over four years and suggested that bronchial Candida colonization is an independent factor for pneumonia and the risk is greatest for Pseudomonas VAP [10,23]. In our study we found that 37(60.7 %) of GNB were Multi-Drug Resistant (MDR), 20 of them were associated with C. albicans and 17 of them were associated with Non-Albicans Candida (NAC). Hamet, et al. [12] reported similar findings that Candida colonization was an independent risk factor for MDR bacteria isolation.

We found that mortality is higher with Candida colonization in general and particularly in non-albicans + GNB VAP. Hence, it is essential to ensure that appropriate treatment modalities are sought to enhance survival and quality of life of these critically ill patients. Therefore, it is crucial to consider further studies on the possible role of treatment and whether this would result in reduced risk of colonization. Conflicting findings about the role of treatment with antifungal agents have been reported. Nasir, et al. [24] showed that antifungal treatment decreased the risk of Pseudomonas VAP in tracheobronchial Candida Colonized Patients. In another more recent study (the famous CANTREAT study) [25], with antifungal treatment, mortality and length of hospital stay were not reduced. Ong, et al. [26] studied nebulized antifungal treatment as a part of decolonization. They found that nebulized amphotericin B reduces the duration of Candida colonization but failed to improve clinical outcomes [26].

Although selective digestive decontamination in mechanically ventilated patients significantly decreased the colonization rate of GNB and of Candida species as demonstrated in many studies, the role remains unclear and hence not recommended in modern practice [27,28].

STRENGTH AND LIMITATIONS

Strength

The strength of our study comes from its unique design and that the study was positioned at a time period before the patients were placed on antifungal treatment. In addition, those with candidemia were excluded to ensure that mortality due to Candida could be directly estimated among critically ill patients on MV in our hospital.

Limitations

Due to small sample size, we could not include the data on grampositive bacteria + Candida. However, it was not thought that this would enhance the study much more. Another weakness of this study is the absence of histopathologic examination of enrolled patients. However, from a previous study among patients and in post-mortem studies, the incidence of invasive candidiasis were extremely low [29]. Since this study was done on mechanically ventilated patients with Candida colonization with or without GNB VAP, the clinical outcome found in this study population cannot be generalized to other patients including those with other respiratory diseases such as cystic fibrosis who are not mechanically ventilated.

CONCLUSION

There is a high mortality rate with Candida species colonization especially with concomitant GNB VAP. The highest mortality occurred with C. non-albicans. To our knowledge, this is the first study from Saudi Arabia that assessed USCS mortality in mechanically ventilated patients colonized with Candida species + GNB VAP. Further studies are still needed to determine whether treating or decolonizing the respiratory tract with antifungal therapy will decrease GNB VAP, length of hospital stay or mortality.

REFERENCES

- Blot S, Vandewoude K. Management of invasive candidiasis in critically ill patients. Drugs. 2004;64(19):2159-75. doi: 10.2165/00003495-200464190-00002. PMID: 15456333.
- Blot S, Dimopoulos G, Rello J, Vogelaers D. Is Candida really a threat in the ICU? Current Opinion in Critical Care. 2008 Oct;14(5):600-604. doi: 10.1097/ mcc.0b013e32830f1dff.
- Fridkin SK, Jarvis WR. Epidemiology of nosocomial fungal infections. Clin Microbiol Rev. 1996 Oct;9(4):499-511. doi: 10.1128/CMR.9.4.499-511.1996. PMID: 8894349; PMCID: PMC172907.
- Leon C, Ruiz-Santana S, Saavedra P. Usefulness of the "Candida score" for discriminating between Candida colonization and invasive candidiasis in non-neutropenic critically ill patients: a prospective multicenter study. J Crit Care Med 2009;37:1624-1633.
- Laverdiere M, Labbe AC, Restieri C. Susceptibility patterns of Candida species recovered from Canadian intensive care units. J Crit Care. 2007;22:245-250. doi: 10.1016/j.jcrc.2006.10.038.
- Souza RC, Junqueira JC, Rossoni RD, Pereira CA, Munin E, Jorge AO. Comparison of the photodynamic fungicidal efficacy of methylene blue, toluidine blue, malachite green and low-power laser irradiation alone against Candida albicans. Lasers Med Sci. 2010 May;25(3):385-9. doi: 10.1007/ s10103-009-0706-z. Epub 2009 Jul 5. PMID: 19579004.
- Souza RC, Junqueira JC, Rossoni RD, Pereira CA, Munin E, Jorge AO. Comparison of the photodynamic fungicidal efficacy of methylene blue, toluidine blue, malachite green and low-power laser irradiation alone against Candida albicans. Lasers Med Sci. 2010 May;25(3):385-9. doi: 10.1007/ s10103-009-0706-z. Epub 2009 Jul 5. PMID: 19579004.
- Pittet D, Monod M, Suter PM, Frenk E, Auckenthaler R. Candida colonization and subsequent infections in critically ill surgical patients. Ann Surg. 1994 Dec;220(6):751-8. doi: 10.1097/00000658-199412000-00008. PMID: 7986142; PMCID: PMC1234477.
- Rello J, Esandi ME, Díaz E, Mariscal D, Gallego M, Vallès J. The role of Candida sp isolated from bronchoscopic samples in nonneutropenic patients. Chest. 1998 Jul;114(1):146-9. doi: 10.1378/chest.114.1.146. PMID: 9674461.
- Azoulay E, Timsit JF, Tafflet M. Candida colonization of the respiratory tract and subsequent pseudomonas ventilator-associated pneumonia. Chest. 2006;129:110-117. doi: 10.1378/chest.129.1.110
- Delisle MS, Williamson DR, Albert M, Perreault MM, Jiang X, Day AG, Heyland DK. Impact of Candida species on clinical outcomes in patients with suspected ventilator-associated pneumonia. Can Respir J. 2011 May-Jun;18(3):131-6. doi: 10.1155/2011/827692. PMID: 21766075; PMCID: PMC3328877.
- Hamet M, Pavon A, Dalle F, Pechinot A, Prin S, Quenot JP, Charles PE. Candida spp. airway colonization could promote antibiotic-resistant bacteria selection in patients with suspected ventilator-associated pneumonia. Intensive Care Med. 2012 Aug;38(8):1272-9. doi: 10.1007/s00134-012-2584-2. Epub 2012 Jun 15. PMID: 22699790.
- Delisle MS, Williamson DR, Perreault MM, Albert M, Jiang X, Heyland DK. The clinical significance of Candida colonization of respiratory tract secretions in critically ill patients. J Crit Care. 2008;23:11-17.
- Deorukhkar SC, Saini S, Mathew S. Non-albicans Candida Infection: An Emerging Threat. Interdiscip Perspect Infect Dis. 2014;2014:615958. doi: 10.1155/2014/615958. Epub 2014 Oct 22. PMID: 25404942; PMCID: PMC4227454.
- Pongracz J, Benedek K, Juhasz E, Ivan M, Kristof K. In vitro biofilm production of Candida bloodstream isolates: Any association with clinical characteristics? J Med Microbiol. 2016;65:272-277.

American J Anesth Clin Res

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- Sardi JCO, Scorzoni L, Bernardi T, Fusco-Almeida AM, Mendes Giannini MJS. Candida species: current epidemiology, pathogenicity, biofilm formation, natural antifungal products and new therapeutic options. J Med Microbiol. 2013 Jan;62(Pt 1):10-24. doi: 10.1099/jmm.0.045054-0. Epub 2012 Nov 22. PMID: 23180477.
- Marcos-Zambrano LJ, Escribano P, Bouza E, Guinea J. Comparison of the antifungal activity of micafungin and amphotericin B against Candida tropicalis biofilms. J Antimicrob Chemother. 2016 Sep;71(9):2498-501. doi: 10.1093/jac/dkw162. Epub 2016 May 4. PMID: 27147303.
- Peleg AY, Hogan DA, Mylonakis E. Medically important bacterial-fungal interactions. Nat Rev Microbiol. 2010 May;8(5):340-9. doi: 10.1038/ nrmicro2313. Epub 2010 Mar 29. PMID: 20348933.
- Morales DK, Hogan DA. Candida albicans interactions with bacteria in the context of human health and disease. PLoS Pathog. 2010 Apr 29;6(4):e1000886. doi: 10.1371/journal.ppat.1000886. PMID: 20442787; PMCID: PMC2861711.
- 20. Williamson DR, Albert M, Perreault MM, Delisle MS, Muscedere J, Rotstein C, Jiang X, Heyland DK. The relationship between Candida species cultured from the respiratory tract and systemic inflammation in critically ill patients with ventilator-associated pneumonia. Can J Anaesth. 2011 Mar;58(3):275-84. doi: 10.1007/s12630-010-9439-5. Epub 2010 Dec 14. PMID: 21287306.
- Tan X, Chen R, Zhu S. Candida albicans airway colonization facilitates subsequent Acinetobacter baumannii pneumonia in a rat model. Antimicrob Agents Chemother. 2016;60:3348-3354. doi: 10.1128/AAC.02180-15
- 22. Tan X, Zhu S, Yan D, Chen W, Chen R, Zou J, Yan J, Zhang X, Farmakiotis D, Mylonakis E. Candida spp. airway colonization: A potential risk factor for Acinetobacter baumannii ventilator-associated pneumonia. Med Mycol. 2016 Aug 1;54(6):557-66. doi: 10.1093/mmy/myw009. Epub 2016 Mar 21. PMID: 27001670.

- Ader F, Faure K, Guery B, Nseir S. Pseudomonas aeruginosa and Candida albicans interaction in the respiratory tract: from pathophysiology to a therapeutic perspective. Pathol Biol (Paris) 2008;56:164-169.
- 24. Nseir S, Jozefowicz E, Cavestri B. Impact of antifungal treatment on Candida-Pseudomonas interaction: a preliminary retrospective case-control study. Intensive Care Med. 2007; 33: 137-142. doi: 10.1007/s00134-006-0422-0
- 25. Albert M, Williamson D, Muscedere J, Lauzier F, Rotstein C, Kanji S, Jiang X, Hall M, Heyland D. Candida in the respiratory tract secretions of critically ill patients and the impact of antifungal treatment: a randomized placebo controlled pilot trial (CANTREAT study). Intensive Care Med. 2014 Sep;40(9):1313-22. doi: 10.1007/s00134-014-3352-2. Epub 2014 Jul 1. PMID: 24981955.
- 26. Ong DS, Klein Klouwenberg PM, Spitoni C, Bonten MJ, Cremer OL. Nebulised amphotericin B to eradicate Candida colonisation from the respiratory tract in critically ill patients receiving selective digestive decontamination: a cohort study. Crit Care. 2013 Oct 11;17(5):R233. doi: 10.1186/cc13056. PMID: 24119707; PMCID: PMC4056077.
- Ferrer M, Torres A, González J, Puig de la Bellacasa J, el-Ebiary M, Roca M, Gatell JM, Rodriguez-Roisin R. Utility of selective digestive decontamination in mechanically ventilated patients. Ann Intern Med. 1994 Mar 1;120(5):389-95. doi: 10.7326/0003-4819-120-5-199403010-00006. PMID: 8304656.
- Hurley JC. Impact of selective digestive decontamination on respiratory tract Candida among patients with suspected ventilator-associated pneumonia. A meta-analysis. Eur J Clin Microbiol Infect Dis. 2016 Jul;35(7):1121-35. doi: 10.1007/s10096-016-2643-7. Epub 2016 Apr 26. PMID: 27116009.
- Meersseman W, Lagrou K, Spriet I, Maertens J, Verbeken E, Peetermans WE, Van Wijngaerden E. Significance of the isolation of Candida species from airway samples in critically ill patients: a prospective, autopsy study. Intensive Care Med. 2009 Sep;35(9):1526-31. doi: 10.1007/s00134-009-1482-8. Epub 2009 Apr 9. PMID: 19357832.