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**Review Article** 

# Current Insights of *H. Pylori* Infection on Gastric Cancer - 🗟

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# ABSTRACT

The incidence rate of stomach cancer is much higher in some regions, such as Asia, and part of Europe, than the rest of the world. The epidemiological studies showed there may be a strong correlation between *Helicobacter pylori* (*H. pylori*), a type of gastritis-causing bacteria, and the carcinogenesis of gastric cancer. Although the precise mechanism is not fully understood yet, it is generally accepted that *H. pylori* may cause gastric cancer by inflammation, mutation and stimulation of cell proliferation. This review summarizes the mechanisms of how this bacterium may stimulate cell proliferation, cause mutations, thus to trigger stomach cancer.

# **INTRODUCTION**

Stomach cancer, also called gastric cancer, starts in the stomach, and most stomach cancers are derived from "The gland cells in the inner stomach lining". The location of stomach cancer can be classified into the following subsites: "cardia, fundus, body, antrum, pylorus, and lesser or greater curvature" [1]. Diffuse and intestinal gastric cancers are the two major histologic types [1]. Although the incidence rate of stomach cancer has dropped in most countries, the mortality rate still remains high, ranking the third among all the cancers [2].

In the United States, stomach cancer is relatively rare compared to some other countries. Data from National Cancer Institute shows that there is an estimate of 28,000 new cases of stomach cancer in 2017, representing 1.7% of all new cancer incidence; there is an estimate of 10,960 people who will die from stomach cancer in 2017, constituting 1.8% of all cancer deaths. In contrast, stomach cancer is more prevalent in other countries, especially in Asian countries. More than 70% stomach cancer cases were observed in developing countries in 2008 [3]. Consistently, the stomach cancer incidence rates for male in 2012 were 35.4 per 100,000 people in Eastern Asia and 20.3 per 100,000 people in Central and Eastern Europe, whereas they were only 5.5 per 100,000 people in North America and 6.7 per 100,000 people in Australia/ New Zealand [4]. Geographical variations in stomach cancer incidence rates are possibly due to different living habits, such as higher use of alcohol or tobacco and higher prevalence of Helicobacter pylori (H. pylori) infection in less developed countries [4]. Additionally, stomach cancer is more prevalent in men than women, which partially may be due to the disruption of the homeostasis of Androgen Receptor (AR) which is activated by the binding of androgen in men [5].

#### Tipa induces inflammatory response and cell proliferation

H. pylori infection has been recognized as one of the reasons causing stomach cancer, but the mechanisms of how this bacterium works in cancer progress is still not very clear. Recent research has made significant progresses and unveiled this bacterium may contribute to cancer development in multiple ways. H. pylori secretes a protein called tumor necrosis factor-a inducing protein (Tipa), which is a strong inducer of chemokine gene expressions [6,7]. Tipa is a homodimer protein consisting of two cysteine residues in the N-terminal domain that can form disulfide bond, and the formation of the homodimer is critical to the induction of carcinogenic activity of Tipa. Tipa is responsible for inducing the expression of a variety of chemokine genes in stomach cancer cells, among which are genes encoding for Ccl2, Ccl7, Ccl20, Cxcl1, Cxcl2, Cxcl5 and Cxcl10. The chemokine family includes CC chemokines, which have two cysteine residues next to each other, and CXC chemokines, which have another amino acid in between the two cysteine residues. The chemokines induced by Tipa are expressed in gastric epithelial cells and the gastric mucosa. The chemokine proteins are able to induce the migration of various types of immune cells, such as natural killer, dendritic cells, and neutrophils, leading to chronic inflammatory responses in gastric epithelial cells and the consequent stomach cancer. This Tipa-chemokine signaling cascade suggests that Tipa is a major player in the stomach cancer development due to *H. pylori* infection [6,7].

Apart from the chemokine genes induced by Tipa secreted from *H. pylori* that contributes to stomach cancer, Tipa also induces a lot of other genes resulting in gastric carcinogenesis [6]. The expression of interleukin 6 (IL-6),  $\beta$ -catenin, ATP-binding cassette, sub-family B member 4 (Abcb4), Mid-1-related chloride channel 1 (Mclc), and reticuloendotheliosis oncogene (Rel) is induced by Tipa secreted from *H. pylori*. Unlike chemokines inducing a series of inflammatory reaction, these genes are responsible for cell proliferation [6]. Furthermore, the activation of NF- $\kappa$ B, which is a target of Tipa, stimulates the gene expression of chemokines, IL-6, etc. to favor inflammation and cell proliferation [6]. Taken together, Tipa plays an important role in stomach cancer development via both inflammation and inducing cell proliferation.

In addition, Tipa secreted by *H. pylori* stimulates inflammatory response via not only the induction of chemokines, but the induction of Tumor Necrosis Factor- $\alpha$  (TNF $\alpha$ ). TNF $\alpha$  activates NF- $\kappa$ B, which then activates prostaglandin-endoperoxide synthase 2 (COX-2). COX-2 can not only suppress cell apoptosis, but produce prostaglandin E<sub>2</sub>, which induces an inflammatory response [6-8].

Another important aspect of Tipa in terms of carcinogenesis is the fact that Tipa may induce Epithelial-Mesenchymal-Transition (EMT) in gastric epithelial cells [9,10]. Work done by Chen G. et al. and Han Y. et al. showed that Tipa may decrease the expression of E-cadherin and Programmed-Cell-Death-Protein 4 (PDCD4), as well as increase the expression of N-cadherin, vimentin, and TWIST1. The decreased expression of E-cadherin may result in the activation of β-catenin, which could modify the expression of cyclin D1, CD44, c-Myc, etc., facilitating tumor progression [11]. The PDCD4 mainly contributes to the inhibition of transcription and translation of TWIST1 and the inhibition of invasion-related urokinase plasminogen activator receptor expression; as a result, the lower level of PDCD4 triggered by Tipa promotes tumorigenesis [10]. Moreover, the changed expression levels of these markers during EMT may cause cells to lose adhesion and polarity, to increase mobility, to become more stemcell like, to resist to apoptosis, and to degrade extracellular matrix. All these aspects could accelerate tumor malignancy and metastasis in gastric cancer [10]. Additionally, the pathway through which Tipa induces EMT process was determined to be IL-6/STAT3 [12].

## The virulence factor of H. pylori: CagA and VacA

Different strains of *H. pylori* bacteria may possess different virulence factors. The virulence factors may be membrane-associated, secreted or translocated into cytosol of the host cells. Among those factors, cytotoxin-associated gene A (CagA), vacuolating cytotoxin

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(VacA) and outer membrane proteins (OMPs) are directly or indirectly implicated in H. pylori carcinogenesis [13,14]. For example, CagA protein can be directly translocated into B cells where it inhibits apoptosis and promotes cell proliferation [15]. In addition, CagA leads to silencing of the tumor suppressor genes RUNX1, TFF1, and CDH1 by increased methylation in the promoter region. CagA can also activate the expression of the oncogenes EGFR, MET, PI3K, AKT, and CTNNB1 (which codes for  $\beta$ -catenin) [16]. Furthermore, H. pylori infection with the presence of CagA and VacA has a synergistic effect on the development of gastric cancer with IL-1β gene polymorphisms, and the highest prevalence of severe gastric abnormalities are found in patients with both host and bacterial high-risk genotypes (cagA(+)/vacAs1(+)/IL-1β-511T) [17]. In a more recent study on stem cell-derived gastric organoids, CagA activated the c-Met receptor and resulted in a rapid induction of epithelial cell proliferation [18]. CagA could also activate the Wnt/β-catenin signaling pathway, leading to the transcription of oncogenes [10]. Moreover, CagA may induce EMT process, which partially depends on the regulation of PDCD4 [10]. The expression level of CagA may be elevated by high salt diet [19].

In addition to the carcinogenesis of CagA, VacA of H. pylori also plays an essential role in both induction of apoptosis in gastric epithelial cells and vacuolating cytotoxicity [20,21]. Studies by Kuck Dirk et al. demonstrated that VacA may activate the CD95 receptor and ligand system to induce apoptosis [21,22]. Moreover, VacA can form a hexameric anion-selective channel in cell membrane at an acidic pH, through which the toxin gets transmitted into cytosol, and the toxin may interfere with a substance that controls the membrane trafficking of endosomes and lysosomes. Mucosal damage and the consequent gastric tumorigenesis can be attributed to both factors of VacA of H. pylori infection [21].

# H. pylori upregulates reactive species in host individuals

The infection of H. pylori may increase the formation of oxy- and nitro- radical species in gastric mucosa, such as Myeloperoxidase (MPO), inducible Nitric Oxide Synthase (iNOS), and NADPH oxidase [9]. The expression of these reactive species is derived from activated macrophages, neutrophils, or monocytes [23,24]. The longterm effect of the radical species may include inflammation, tissue damage, and carcinogenesis [23]. It is worth noting that Reactive Oxygen Species (ROS) and Nitric Oxide (NO), which is a product of iNOS, can cause DNA damage as well. During NO synthesis, DNA deamination may occur, and thus mutations of gastric epithelial cells could happen. Additionally, the release of ROS and NO may result in DNA methylation in the CpG Island in promoter, causing genetic alterations and thus carcinogenesis [9].

#### H. pylori downregulates host DNA repair mechanisms

Another negative impact H. pylori may have on the stomach tissue is that the bacteria may downregulate DNA repair mechanisms; as a result, host genome becomes increasingly unstable, allowing potential mutations in the tumor suppressor genes and oncogenes to occur [25,26]. The impacts of H. pylori include reduction in mismatched repair, base excision repair, and double-strand break repair [26]. The reduced stability of host genome and downregulation of DNA repair contribute to carcinogenesis through increasing the host cells' possibility of mutations and their maintenance. The reduced ability of DNA repair mechanisms render the host cells less protected from exogenous insults.

# **TREATMENT OPTIONS**

H. pylori infection has been associated with various gastric problems, including gastric cancer. However, treatment of H. pylori remains a challenge, due to the rising prevalence of antimicrobial resistance, mainly to clarithromycin, efficacy of standard triple therapies has declined to unacceptably low levels in most parts of the world. Novel regimens, such as bismuth quadruple, concomitant, have been shown to improve the therapeutic outcome against antibiotic-resistant H. pylori strains [27].

In addition, a vaccine would overcome these drawbacks, but currently there is not any H. pylori vaccine licensed. In seeking alternative treatment, the use of probiotics has been proposed in order to optimize the eradication rates. Several clinical trials indicated that administration of probiotics can reduce the side effects of H. pylori eradication treatment by antibiotics, increasing tolerability and often increasing the overall efficacy [28].

Gastrin is responsible for the secretion of gastric acid, and hyperacidity is a contributing factor to H. pylori infection. Curcumin is an anticancer agent, which is naturally from the root of Curcuma longa and exerts the anti-H. pylori effect by inhibiting the acid secretion in stomach. Inhibiting the level of gastrin secretion, curcumin significantly raises gastric pH in vivo and prevents H. pylori infection [29].

Another anticancer agent is evodiamine, which is an alkaloid compound that naturally occurs in Evodia rutaecarpa. It may induce autophagy and apoptosis of gastric cancer cells. During the process of autophagy, auto phagosomes are formed to degrade tumor cells. On the other hand, the pro-survival gene expression of Bcl-2 in gastric cancer cells is inhibited by evodiamine, while the pro-apoptotic gene expression of Bax is activated [30].

# PREVENTION

To prevent stomach cancer caused by H. pylori infection, antiinflammatory agents and anti-oxidants could be applied during the course of H. pylori treatment [31]. Nonsteroidal anti-inflammatory drugs can be promising to eradicate inflammatory response due to H. pylori infection and thus prevent from stomach cancer. Anti-oxidants can be applied with dietary supplementation, such as ascorbic acid and β-carotene, since they can "induce regression of precancerous lesions in patients with intestinal metaplasia and/or multifocal atrophic gastritis" [31]. A clinical trial demonstrated that the eradication of H. pylori indeed slowed down the stomach cancer progression [31].

Moreover, the National Cancer Institute suggests that avoiding adverse lifestyle may lower the risk of getting stomach cancer, such as smoking cessation and healthy dietary. Low intake of salt, high intake of fresh fruits and vegetables that contain vitamin C, and high intake of whole-grain cereals, carotenoids, allium compounds, and green tea may all decrease the risk of getting stomach cancer.

# REFERENCES

- 1. Jim MA, Pinheiro PS, Carreira H, Espey DK, Wiggins CL ,Weir HK. Stomach cancer survival in the United States by race and stage (2001-2009): findings from the CONCORD-2 study. Cancer. 2017; 24: 4994-5013. https://goo.gl/ ak5e3b
- 2. Lan Mi, Xin Ji, Jiafu Ji. Prognostic biomarker in advanced gastric cancer. Transl Gastrointest Cancer, 2016; 5; 16-29, https://goo.gl/XwTVRW
- 3. Ferlay J, Shin H, Bray F, Forman D, Mathers C, Parkin DM. Estimates of worldwide burden of cancer in 2008: GLOBOCAN 2008. Int J Cancer. 2010; 127: 2893-2917. https://goo.gl/waf2et

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- Torre LA, Bray F, Siegel RL, Ferlay J, Lortet Tieulent J, Jemal A. Global cancer statistics, 2012. CA Cancer J Clin. 2015; 65: 87-108. https://goo.gl/ s2Bff6
- Tian, Y, Wan H, Lin Y, Xie X, Li Z, Tan G. Androgen receptor may be responsible for gender disparity in gastric cancer. Med Hypotheses. 2013; 80: 672-674. https://goo.gl/TXV9nz
- Kuzuhara T, Suganuma M, Kurusu M, Fujiki H. Helicobacter pylori-secreting protein Tipα is a potent inducer of chemokine gene expressions in stomach cancer cells. J Cancer Res Clin Oncol. 2007; 133: 287-296. https://goo.gl/ fz4rqL
- Tang CL, Hao B, Zhang GX, Shi RH, Cheng WF. Helicobacter pylori tumor necrosis factor-α inducing protein promotes cytokine expression via nuclear factor-κB. World J Gastroenterol. 2013; 19: 399-403. https://goo.gl/R5HMuo
- Watanabe T, Takahashi A, Suzuki K, Kurusu-Kanno M, Yamaguchi K, Fujiki H. Epithelial-mesenchymal transition in human gastric cancer cell lines induced by TNF-α-inducing protein of Helicobacter pylori. Int J Cancer. 2014; 134: 2373-2382. https://goo.gl/jSDfmu
- Elfvin A, Edebo A, Hallersund P, Casselbrant A, Fändriks L. Oxidative and nitrosative stress enzymes in relation to nitrotyrosine in Helicobacter pyloriinfected humans. World J Gastrointest Pathophysiol. 2014; 5: 373-379. https://goo.gl/puKrVj
- de Martel C, Ferlay J, Franceschi S, Vignat J, Bray F, Forman D, et al. Global burden of cancers attributable to infections in 2008: a review and synthetic analysis. Lancet Oncol. 2012; 13: 607-615. https://goo.gl/tBEsFn
- Loh JT, Torres VJ, Cover TL. Regulation of Helicobacter pylori cagA expression in response to salt. Cancer Res. 2007; 67: 4709-4715. https:// goo.gl/xtYZoQ
- Chey WD, Wong BC, Practice Parameters Committee of the American College of Gastroenterology. American College of Gastroenterology guideline on the management of Helicobacter pylori infection. Am J Gastroenterol. 2007; 102: 1808-1825. https://goo.gl/5eSbX3
- Ajani JA, Lee J, Sano T, Janjigian YY, Fan D, Song S. Gastric adenocarcinoma. Nat Rev Dis Primers. 2017; 3: 17036. https://goo.gl/VTn4QC
- Cover TL, Peek RM Jr. Diet, microbial virulence, and Helicobacter pyloriinduced gastric cancer. Gut Microbes. 2013; 4: 482-493. https://goo.gl/ b5URPu
- McCracken KW, Catá EM, Crawford CM, Sinagoga KL, Schumacher M, Rockich BE et al. Nature. 2014; 516: 400-404. https://goo.gl/J3PXqY
- 16. Chatre L, Fernandes J, Michel V, Fiette L, Avé P, Arena et al. Helicobactoer pylori targets mitochondrial import and components of mitochondrial DNA replication machinery through an alternative VacA-dependent and a VacAindependent mechanisms. Sci Rep. 2017; 7: 15901. https://goo.gl/oe2EF7
- Chmiela M, Karwowska Z, Gonciarz W, Allushi B, Staczek P. Host pathogen interactions in Helicobacter pylori related gastric cancer. World Journal of Gastroenterology. 2017; 23: 1521-1540. https://goo.gl/r9CGnh

- Papastergiou V, Georgopoulos SD, Karatapanis S. Treatment of Helicobacter pylori infection: Past, present and future. World J Gastrointest Pathophysiol. 2014; 5: 392-399. https://goo.gl/yofl\_g5
- Ruggiero P. Use of probiotics in the fight against Helicobacter pylori. World J Gastrointest Pathophysiol. 2014; 5:384-391. https://goo.gl/fcD8s7
- Zhou S, Yao D, Guo L, Teng L. Curcumin suppresses gastric cancer by inhibiting gastrin-mediated acid secretion. FEBS Open Bio. 2017; 7: 1078-1084. https://goo.gl/P6LiC3
- 21. Wu M, Chen C, Lin J. Host-environment interactions: their impact on progression from gastric inflammation to carcinogenesis and on development of new approaches to prevent and treat gastric cancer. Cancer Epidemiol Biomarkers Prev. 2005; 14: 1878-1882. https://goo.gl/wQPJHr
- 22. Hong JB, Zuo W, Wang AJ, Lu NH. Helicobacter pylori Infection Synergistic with IL-1β Gene Polymorphisms Potentially Contributes to the Carcinogenesis of Gastric Cancer. Int J Med Sci. 2016; 13: 298-303. https://goo.gl/45JGB4
- 23. Chen G, Tang N, Wang C, Xiao L, Yu M, Zhao L, et al. TNF-α-inducing protein of Helicobacter pylori induces epithelial-mesenchymal transition (EMT) in gastric cancer cells through activation of IL-6/STAT3 signaling pathway. Biochem Biophys Res Commun. 2017; 484: 311-317. https://goo.gl/UdgPtW
- Nardone G, Rocco A, Malfertheiner P. Review article: helicobacter pylori and molecular events in precancerous gastric lesions. Aliment Pharmacol Ther. 2004; 20: 261-270. https://goo.gl/TsuW7G
- Goto T, Haruma K, Kitadai Y, Ito M, Yoshihara M, Sumii K, et al. Clin Cancer Res. 1999; 5: 1411-1415. https://goo.gl/1kQQs8
- 26. Yu H, Zeng J, Liang X, Wang W, Zhou Y, Sun Y, et al. Helicobacter pylori promotes epithelial-mesenchymal transition in gastric cancer by downregulating programmed cell death protein 4 (PDCD4). PLoS ONE. 2014; 9: 105306. https://goo.gl/r98fkB
- Huang L, Wu RL, Xu AM. Epithelial-mesenchymal transition in gastric cancer. Am J Transl Res. 2015; 7: 2141-2158. https://goo.gl/6tb16M
- Xia HH, Talley NJ. Apoptosis in gastric epithelium induced by Helicobacter pylori infection: implications in gastric carcinogenesis. Am J Gastroenterol. 2001; 96: 16-26. https://goo.gl/hwKVgB
- Kuck D, Kolmerer B, Iking-Konert C, Krammer PH, Stremmel W, Rudi J. Vacuolating cytotoxin of Helicobacter pylori induces apoptosis in the human gastric epithelial cell line AGS. Infect Immun. 2001; 69: 5080-5087. https:// goo.gl/moCZpR
- Rudi J, Kuck D, Strand S, von Herbay A, Mariani SM, Krammer PH, et al. Involvement of the CD95 (APO-1/Fas) receptor and ligand system in Helicobacter pylori-induced gastric epithelial apoptosis. J Clin Invest. 1998; 102: 1506-1514. https://goo.gl/cMXbhW
- Rasul A, Yu B, Zhong L, Khan M, Yang H, Ma T. Cytotoxic effect of evodiamine in SGC-7901 human gastric adenocarcinoma cells via simultaneous induction of apoptosis and autophagy. Oncol Rep. 2012; 27: 1481-1487. https://goo.gl/ zaStso